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Scheme 1. Peloruside A (1): Structure and Strategy. PG = protecting group.

Peloruside A (1) is a potent cytotoxin and induces apoptosis in cultured mammalian cells.[2] Of potential clinical relevance is the recent observation that 1 affects microtubule dynamics in a manner similar to paclitaxel (Taxol).[2b] The relative stereochemistry of 1 was assigned based on extensive data gathered from a variety of high-field NMR experiments, although the absolute configuration remained to be solved. In addition to the presence of an array of ten stereogenic centers and a Z-configured trisubstituted olefin, we anticipated that one of the main challenges of a peloruside synthetic venture would be to find an efficient solution to the sterically crowded C8-C11 unit. In the solution-phase structure of 1, this region adopts a conformation determined by optimal minimization of syn and gauche pentane interactions with potential intramolecular hydrogen bonding between the C9 and C11 hydroxy groups (A, Scheme 1). On a synthetic pathway, the introduction of the C8, C9, and C11 hydroxy functionality will by necessity introduce additional syn pentane and dipoledipole interactions depending on the protection scheme used (B, Scheme 1). Herein, we communicate a solution to these synthetic challenges that culminates in the first total synthesis of (-)-peloruside A (1), and in doing so, document the absolute configuration of the natural isolate (opposite to the one shown in Scheme 1).[3]

Strategically, we opted for a late-stage aldol coupling between a fully functionalized aldehyde I with methyl ketone II. This approach is driven by the perception that access to diastereomeric seco-acids, through reduction of a common C15 ketone, will increase the odds for a successful macrocyclization by accessing two mechanistically distinct pathways, namely acylative or invertive macrolactonization. A concise, enantioselective synthesis of fragment II is outlined in Scheme 2. The known homoallylic alcohol 2, prepared in enantiopure form according to an elegant procedure described by Hoveyda et al., [4] was acylated with methacryloyl chloride followed by ring-closing olefin metathesis with Grubb's second generation catalyst 4.^[5] The resulting lactone 5 then provides a valuable entry to the Z-trisubstituted enone

Macrolide Total Synthesis



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Recently, we became interested in peloruside A (1, Scheme 1), a marine metabolite isolated from sponge specimens of the species Mycale by Northcote and co-workers.[1]

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Scheme 2. a) $CH_2CMeC(O)Cl$, iPr_2NEt , DMAP, CH_2Cl_2 (75%); b) 10 mol% **4**, CH_2Cl_2 (0.0025 M), reflux, 17 h (50–70% + 20% dimer derived from **3**); c) MeLi, THF, -78 °C; or Me_3SiCH_2Li , pentane, -78 °C; d) TBSCl, imidazole, DMAP, DMF (52–63% from **5**). DMAP = 4-dimethylaminopyridine, TBS = tert-butyldimethylsilyl.

6 by treatment with methyllithium and silylation of the primary alcohol.

Our approach towards the densely functionalized tetrahydropyranyl fragment is shown in Scheme 3. Addition of aldehyde 9 (derived from $7^{[6]}$ as shown) to (Z)-alkoxyallylborane 10 at low temperature produced the desired homoallylic alcohol diastereomer 11 (> 10:1).^[7] The relative 1,3-syn stereochemical relationship was demonstrated by 13 C NMR analysis of the acetonide derivative 13 (resonances for the isopropylidene carbon atoms at $\delta = 20.0$, 30.3, and 98.8 ppm).^[8] Compound 11 was advanced to aldehyde 15 by methylation (\rightarrow 12), oxidative debenzylation (DDQ), and oxidation of the resulting primary alcohol. Addition of aldehyde 15 to a cold solution of the lithium enolate derived from 14 and Dess–Martin oxidation^[9] of the resulting aldol product fashioned β -diketone 16. Transformation to dihydropyranone 17 ensued by stirring 16 in an acidic toluene

Scheme 3. a) TESOTf, 2,6-lutidine, CH_2Cl_2 (97%); b) cat. OsO_4 , NMO, acetone/ H_2O ; c) $Pb(OAc)_4$, pyridine, CH_2Cl_2 (80% from 8); d) 10, sBuLi, THF, -78°C, 15 min, then (+)-lpc₂BOMe, -78°C, 1 h, $-78 \rightarrow 0$ °C, 1.5 h, then 9, -95°C, 3 h, slowly warm to RT; 30% H_2O_2 , NaOH, 16 h (91%); e) NaH, Mel, DMF, -5°C (89%); f) PTSA, MeOH, 20 min, remove solvent, then $Me_2C(OMe)_2$ (44%); g) DDQ, CH_2Cl_2/H_2O , 0°C (88%); h) $py \cdot SO_3$, Et_3N , DMSO, CH_2Cl_2 , 0°C (87%); i) 14, LDA, THF, -78°C, then 15, -78°C (94%); j) Dess-Martin periodinane, CH_2Cl_2 , -10°C (81% based on recovered starting material); k) PTSA, PhMe, RT (92%); l) $NaBH_4$, $CeCl_3 \cdot 7H_2O$, MeOH, -30°C; m) mCPBA, $NaHCO_3$, $CH_2Cl_2/MeOH$, 0°C (72%, two steps); n) tBuOK, Mel, THF, 0°C; o) TESOTf, 2,6-lutidine, CH_2Cl_2 (75% from 19); p) cat. OsO_4 , OsO_4 , OsO_4 , OsO_5 , OsO_6 ,

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solution. It proved critical to perform the subsequent Luche^[10] reduction (\rightarrow **18**) at -30 °C to prevent decomposition. Most satisfactorily, hydroxy-directed epoxidation of crude 18 followed by in situ methanolysis of the incipient glycal-epoxide produced one single glycoside, 19, in high yield (72% from 17).[11] Sequential methylation (equatorial OH)/silylation (axial OH) of the α -diol (\rightarrow 20), oxidative transformation of the double bond to a C1 carboxylic acid, and diazomethane treatment furnished methyl ester 21. Access to the aldehyde 22 by hydrogenolytic removal of the benzyl ether and oxidation set the stage for progression through C11-C12 bond formation. Concerns related to the feasibility of this critical event[12] seemed justified when various attempts to add carbon nucleophiles to the sterically demanding aldehyde 22 failed.[13] Ultimately, a highly efficient allyl transfer was achieved with allyldiethylborane. Remarkably, a single alcohol diastereomer, 23, was produced with C11 configuration ascertained by NOE analysis of spirocyclic ketal 24 and advancement of 23 to peloruside A (see below).[14]

With fragments 6 and 25 (derived from 24 as shown) at hand, their union and completion of the peloruside macrocycle seemed an attainable goal, yet unexpected surprises lay ahead (Scheme 4). Mukaiyama-type aldol reaction of aldehyde 25 with the enol silane derived from methyl ketone 6 afforded almost exclusively (14:1) the compound that was

Scheme 4. a) Enolsilane derived from **6** (TMSOTf, Et₃N, CH₂Cl₂, $-10\,^{\circ}$ C, 25 min, aqueous extraction), **25**, CH₂Cl₂, $-78\,^{\circ}$ C, add BF₃·Et₂O, 2 h (80%, 14:1 ratio); b) Me₃OBF₄, 1,8-bis (dimethylamino) naphthalene, CH₂Cl₂, RT (92%); c) (S)-B-Me-CBS (20 equiv), BH₃·SMe₂ (7 equiv), CH₂Cl₂, $-30\,^{\circ}$ C, 1 h, \rightarrow RT (4 h), add MeOH (83%, 13:1 ratio); d) 0.3 N aq. LiOH, THF, RT (quant.); e) PPh₃, DIAD, THF (0.05 M), RT, 15 min, add **28** (0.003 M in THF) through syringe pump over 2 h, 1 h at RT (40–50%); f) 48% aq. HF, MeCN/H₂O, RT, 20 h (88%). TMS = trimethylsilyl, CBS = catalyst named after Corey, Bakshi, and Shibata, DIAD = diisopropylazodicarboxylate.

assumed to be the expected 1,3-anti aldol product based on extensive literature precedent. [15] Evidence provided below, however, indicates that aldehyde **25** entered this reaction with an unprecedented bias for the formation of the 1,3-syn β -hydroxy ketone **26** instead (80% yield). Initially unaware of this stereochemical outcome, we continued with methyl ether formation (\rightarrow 27), CBS reduction, [16] and ester hydrolysis to reach seco-acid **28**. Slow addition of this compound to a premixed solution of PPh₃ and diisopropylazodicarboxylate instigated formation of macrolactone **29** in 40–50% yield. [17] We were able to deduce the stereochemistry of **29** based on a series of NOE correlations that locate H11, H13, and H15 on the same upper side of the macrolactone ring in agreement with the assigned C13(S) configuration (Figure 1). At this

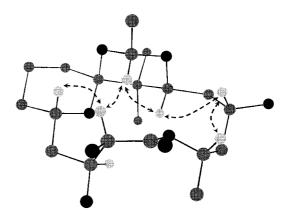


Figure 1. Chem3D representation of 29 (C in dark gray, H in light gray, and O in black). Important NOEs are illustrated with dashed arrows.

point, we were left with the obvious challenge of correcting the stereochemistry at C13. Various attempts to remove the (2-naphthyl)methylidene acetal failed. Undeterred, we embraced the opportunity to explore a more attractive avenue that would eliminate this protecting group problem altogether, that is, we decided to advance materials with a free C11 alcohol through the remainder of the synthesis.

As illustrated in Scheme 5, β-hydroxy aldehyde 31 (derived from 23 as shown) was a viable partner for coupling with the enolborinate derived from 6. Most gratifyingly, a separable 2:1 mixture of C13 epimers (32a/32b) was generated in 87% yield. This rather unselective reaction was welcomed when contrasted with the situation described above (cf. 25 + 6 \rightarrow 26). At this stage, we left the stereochemical assignment unresolved, and advanced both isomers 32 a/32 b individually. Surprisingly, methylation of these compounds occurred with concomitant hydrolysis to the C9 hemiketals 33 a/33 b. We did not reprotect these materials but instead positioned ourselves to explore, in line with our original conception, two mechanistically distinct macrocyclization pathways. Intramolecular acylation through an active ester intermediate required access to the C15(R)-configured secoacids 35 a/35b, while Mitsunobu-type lactonization would initiate from the C15-epimeric relatives 34a/34b. These requirements were best fulfilled by asymmetric reduction of enones 33 a/33 b by using (S)- or (R)-B-Me-CBS-oxazabor-

Scheme 5. a) Cat. OsO₄, NMO, acetone/H₂O; b) Pb(OAc)₄, pyridine, CH₂Cl₂ (95% from 23); c) 6, iPr₂NEt, Et₂BOTf, CH₂Cl₂, -78°C, 15 min, -30°C, 45 min, -78°C, add 31, -78°C, 2 h (87%, 2:1 ratio); d) Me₃OBF₄ (20 equiv), 2,6-di-tert-butyl-4-methylpyridine, CH₂Cl₂, RT (85%); e) 33 a or 33 b, (R)- or (S)-B-Me-CBS (20 equiv), BH₃·SMe₂ (7 equiv), CH₂Cl₂, -30°C, 1 h, \rightarrow RT (4 h), add MeOH (34a: 80%, 34b: 80%, 35a: 94%, 35b: 80%); f) 0.3 N aq. LiOH, THF, RT (quant.); g) PPh₃, DIAD, THF (0.05 M), RT, 15 min, add seco-acid (0.003 M in THF) through syringe pump over 2 h, then 1 h at 0°C (36: 47% from 34b or 35b, 52% from ≈1:1 mixture of 34b:35b, 38: 69% from 35a); h) 4 N HCl, THF, RT, 3 h (37: 65%, 1: 65%).

olidine and $H_3B\cdot SMe_2$ (80–94% yield, ratios > 12:1).^[16] Note that no concomitant reduction of the C9 hemiketal was observed under these conditions! The corresponding carboxylic acids **34a,b/35a,b** were liberated by saponification.^[19]

Since the C15 configuration could not be unambiguously inferred from the Corey model, [16] we explored both epimers **34b** and **35b** as substrates for a Mitsunobu-type macrolactonization, Remarkably, the same lactone **36** was produced from either **34b**, **35b**, or an equimolar mixture of both. [20] This lactone was later shown to have a C13 epimeric relationship to peloruside A (see below). [21] To the best of our knowledge, this represents the first documented case of a configuration-dependent mechanistic switch for a Mitsunobu lactonization. [22] We speculate that geometrical/conformational constraints preclude the formation of C15 epimeric lactones [23] and enforce the cyclization of substrate **35b** via an acyloxy-phosphonium intermediate (retention) and **34b** via the familiar alkoxyphosphonium intermediate (inversion). [24,25]

The structure of **36** was assigned retrospectively,^[21] that is, after the completion of peloruside A (**1**) from precursor **35a**. Initial evidence for a C13 epimeric configuration surfaced, however, when the ¹H NMR data of the deprotected lactone

37 (aq. HCl/THF) did not match those reported for the natural product. By inference, we realized peloruside A would fortuitously derive from the major aldol diastereomer 32a. In the event, Mitsunobu lactonization of 35a (69%) followed by simultaneous cleavage of the MOM and silyl protecting groups in lactone 38 (65%), did produce a pure compound (1) with spectroscopic properties (1H and ¹³C NMR, IR, HRMS) identical to those of the natural isolate.[1] However, the optical rotation of synthetic peloruside A (1) was of opposite sign ($[\alpha]_D^{23} = -16$; c = 0.12 in CH_2Cl_2) to the one reported for the natural product ($[\alpha]_D^{20} = +$ 16; c = 0.30 in CH₂Cl₂).^[1] Also, synthetic **1** (up to 10 μм) had no effect on the growth of cultured human tumor cell lines (SK-MEL-5, HeLa).[26] Based on these results, we assign the absolute configuration of natural (+)-peloruside A as 2S, 3R, 5R, 7R, 8R, 9R, 11S, 13S, 15S, 18R.

In summary, we accomplished the first total synthesis of (—)-peloruside A and documented the absolute configuration of the dextrorotatory natural product. The advancement of minimally protected intermediates merits recapitulation, and was key to a successful endgame. A particularly noteworthy configuration-dependent mechanistic switch was observed

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when epimeric seco-acids **34b/35b** were subjected to typical Mitsunobu lactonization conditions. Current efforts focus on gaining additional insights into the mechanistic aspects of this reaction, and the synthesis of naturally configured peloruside and tailored derivatives for structure–function studies.

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Keywords: antitumor agents · configuration determination · cyclization · natural products · total synthesis

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- [20] No diastereomeric or transposed (allylic) macrolactones were detected by NMR analysis of the nonpurified mixture.
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